

- [0339] Kobayashi K et al, Cytotoxic effects of benzbromarone and its 1'-hydroxy metabolite in human hepatocarcinoma FLC4 cells cultured on micro-space cell culture plates. *Drug Metab Pharmacokinet.* 2013; 28(3):265-8.
- [0340] Korff T et al. Integration of endothelial cells in multicellular spheroids prevents apoptosis and induces differentiation. *J. Cell Biol.* 143: 1341-1352 (1998).
- [0341] Lyons A B. Analysing cell division in vivo and in vitro using flow cytometric measurement of CFSE dye dilution. *J Immunol Methods.* 2000 Sep. 21; 243(1-2): 147-54.
- [0342] Mathias R A et al, Isolation of extracellular membranous vesicles for proteomic analysis. *Methods Mol Biol.* 2009; 528:227-42.
- [0343] Morales P et al, Selective, nontoxic CB(2) cannabinoid o-quinone with in vivo activity against triple-negative breast cancer. *J Med Chem.* 2015 Mar. 12; 58(5): 2256-64. doi: 10.1021/acs.jmedchem.5b00078. Epub 2015 Feb. 20.
- [0344] Morton J J et al, Humanized Mouse Xenograft Models: Narrowing the Tumor-Microenvironment Gap. *Cancer Res.* 2016 Nov. 1; 76(21):6153-6158. Epub 2016 Sep. 1.
- [0345] Nakamura et al., Evaluation of drug toxicity with hepatocytes cultured in a micro-space cell culture system. *J Biosci Bioeng.* 2011 January; 111(1):78-84.
- [0346] Neve et al. "A Collection of Breast Cancer Cell Lines for the Study of Functionally Distinct Cancer Subtypes." *Cancer cell* 10.6 (2006): 515-527
- [0347] Oh WK, Neoadjuvant therapy before radical prostatectomy in high-risk localized prostate cancer: defining appropriate endpoints. *Urol Oncol.* 2003 May-June; 21(3):229-34.
- [0348] Perche F et al, Cancer cell spheroids as a model to evaluate chemotherapy protocols. *Cancer Biology & Therapy* 13:12, 1205-1213, 2012.
- [0349] Phung Y T et al, Rapid Generation of In Vitro Multicellular Spheroids for the Study of Monoclonal Antibody Therapy, *J Canc* 2: 507-514, 2011.
- [0350] Rahman et al. "TRAIL Induces Apoptosis in Triple-Negative Breast Cancer Cells with a Mesenchymal Phenotype." *Breast cancer research and treatment* 113.2 (2009): 217-230.
- [0351] Ramsey S D et al, Integrating comparative effectiveness design elements and endpoints into a phase III, randomized clinical trial (SWOG S1007) evaluating oncotypedX-guided management for women with breast cancer involving lymph nodes. *Contemp Clin Trials.* 2013 January; 34(1):1-9.
- [0352] Reich, M., et al. (2006). "GenePattern 2.0." *Nat Genet* 38(5): 500-501.
- [0353] Rocha N S et al, (2002) Effects of fasting and intermittent fasting on rat hepatocarcinogenesis induced by diethylnitrosamine. *Teratog Carcinog Mutagen.* 22(2): 129-138.
- [0354] Roux S et al, CD4<sup>+</sup>CD25<sup>+</sup> Tregs control the TRAIL-dependent cytotoxicity of tumor-infiltrating DCs in rodent models of colon cancer. *J Clin Invest.* 2008 November; 118(11):3751-61.
- [0355] Ruggeri B A et al, Animal models of disease: pre-clinical animal models of cancer and their applications and utility in drug discovery. *Biochem Pharmacol.* 2014 Jan. 1; 87(1):150-61.
- [0356] Walker J D et al, Oncolytic herpes simplex virus 1 encoding 15-prostaglandin dehydrogenase mitigates immune suppression and reduces ectopic primary and metastatic breast cancer in mice. *J Virol.* 2011 July; 85(14):7363-71.
- [0357] Yang S et al, Mouse models for tumor metastasis. *Methods Mol Biol.* 2012; 928:221-8.
- [0358] Zhang C H et al, Design, Synthesis, and Structure-Activity Relationship Studies of 3-(Phenylethynyl)-1H-pyrazolo[3,4-d]pyrimidin-4-amine Derivatives as a New Class of Src Inhibitors with Potent Activities in Models of Triple Negative Breast Cancer. *J Med Chem.* 2015 May 14; 58(9):3957-74. Epub 2015 Apr. 16.
- [0359] Zhang F et al, ING5 inhibits cancer aggressiveness via preventing EMT and is a potential prognostic biomarker for lung cancer. *Oncotarget.* 2015 Jun. 30; 6(18): 16239-52.
- [0360] Zhang Y et al, Real-Time GFP Intravital Imaging of the Differences in Cellular and Angiogenic Behavior of Subcutaneous and Orthotopic Nude-Mouse Models of Human PC-3 Prostate Cancer. *J Cell Biochem.* 2016 November; 117(11):2546-51.
1. A method of treating a cancer in a subject in need thereof, the method comprising administering to the subject adherent stromal cells (ASC), thereby treating a cancer in a subject.
  - 2-6. (canceled)
  7. The method of claim 1, wherein said ASC have been obtained from a three-dimensional (3D) culture.
  8. The method of claim 7, wherein said 3D culture utilizes a medium whose composition is not varied over the course of said 3D culture.
  9. The method of claim 7, whereby a wherein one or more pro-inflammatory cytokines are added to an incubation medium of said 3D culture.
  10. The method of claim 9, wherein said 3D culture comprises: (a) incubating ASC in a 3D culture apparatus in a first growth medium, wherein no pro-inflammatory cytokines have been added to said first growth medium; and (b) subsequently incubating said ASC in a 3D culture apparatus in a second growth medium, wherein said one or more pro-inflammatory cytokines have been added to said second growth medium.
  - 11-12. (canceled)
  13. The method of claim 9, wherein said one or more pro-inflammatory cytokines comprise Tumor Necrosis Factor alpha (TNF-alpha).
  14. The method of claim 9, wherein said one or more pro-inflammatory cytokines comprise Interferon-Gamma (IFN-gamma).
  15. (canceled)
  16. The method of claim 7, wherein said 3D culture is performed in an apparatus that comprises a 3D bioreactor.
  17. The method of claim 7, wherein said 3D culture is performed in an apparatus that comprises a fibrous bed matrix, said fibrous bed matrix comprising a synthetic adherent material.
  - 18-22. (canceled)
  23. The method of claim 7, further comprising the subsequent step of harvesting said ASC by removing said ASC from an apparatus wherein said 3D culture was performed.
  24. The method of claim 7, wherein said ASC have been incubated in a 2D adherent-cell culture apparatus prior to said 3D culture.